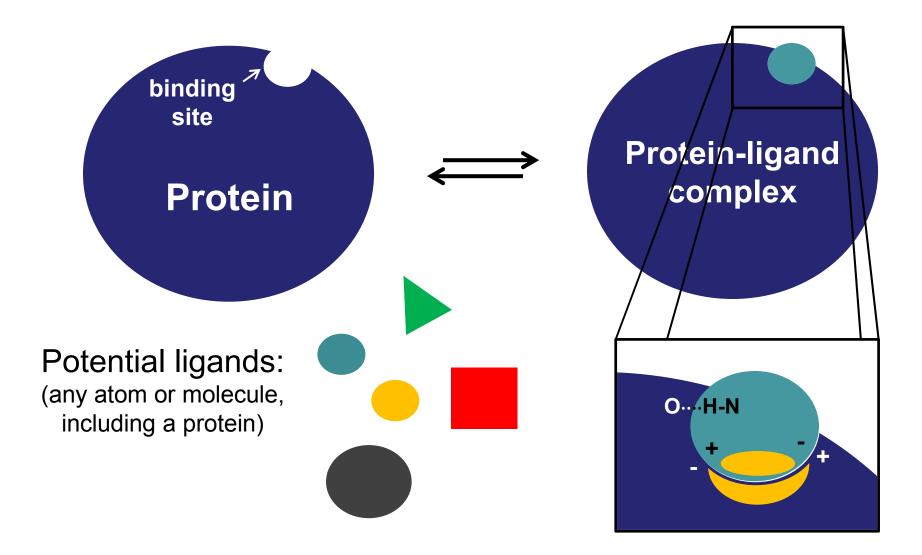
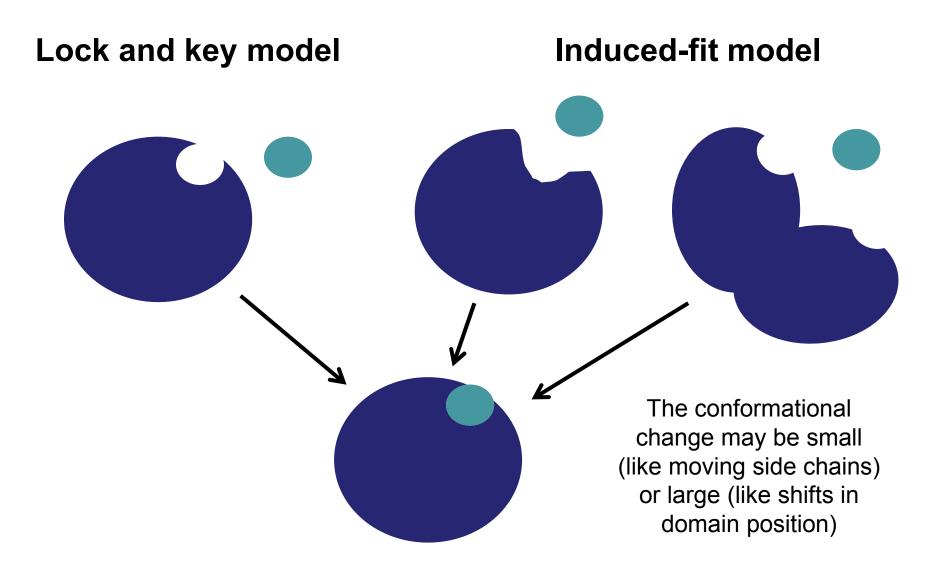
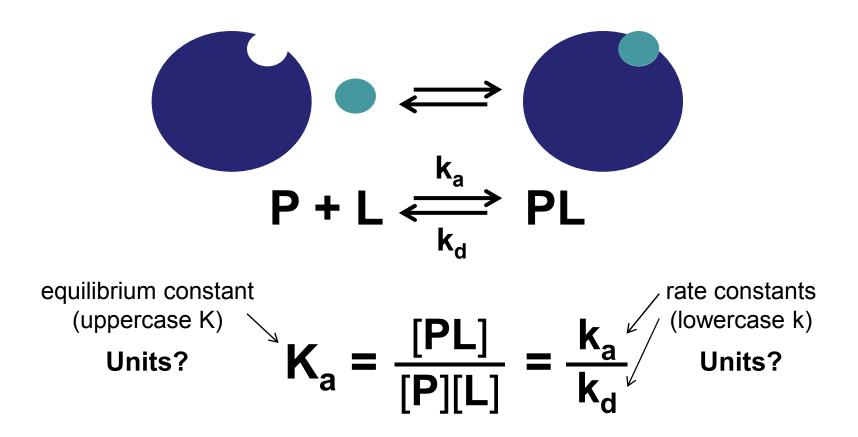
A protein binds a ligand through a specific, reversible interaction



Ligand binding may cause a conformational change in the protein that enhances binding

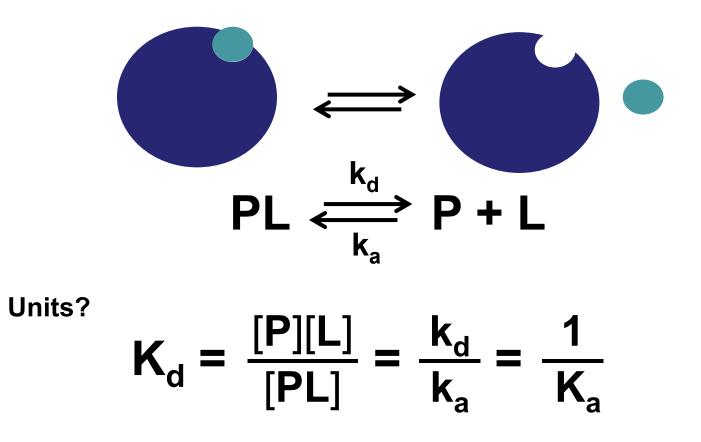


The association constant (K_a) provides a measure of affinity between protein & ligand



Rate constants are proportionality constants, describing the fraction of the pool that reacts in a given amount of time Ex: if $k_d = 0.03 \text{ s}^{-1}$, then 3% of PL dissociates per second

The dissociation constant (K_d) is analogous to the association constant (K_a)



Note: K_a , K_d , k_a , & k_d are constant under set conditions; they can change with changes in temperature, pH, [salt], ...

The fraction of occupied binding sites (θ) is proportional to the ligand concentration

$P + L \iff PL$

When $[L] \gg [PL] + [P]$, [L] is constant (usually true for small ligands in cells)

$$\theta = \frac{binding \ sites \ occupied}{total \ binding \ sites}} = \frac{[PL]}{[PL] + [P]}$$

Substitue in $[PL] = K_a[L][P]$
$$\theta = \frac{K_a[L][P]}{K_a[L][P] + [P]} = \frac{K_a[L]}{K_a[L] + 1} = \frac{[L]}{[L] + \frac{1}{K_a}} = \frac{[L]}{[L] + K_d}$$

The fraction of occupied ligand-binding sites θ depends on [L] and the binding affinity

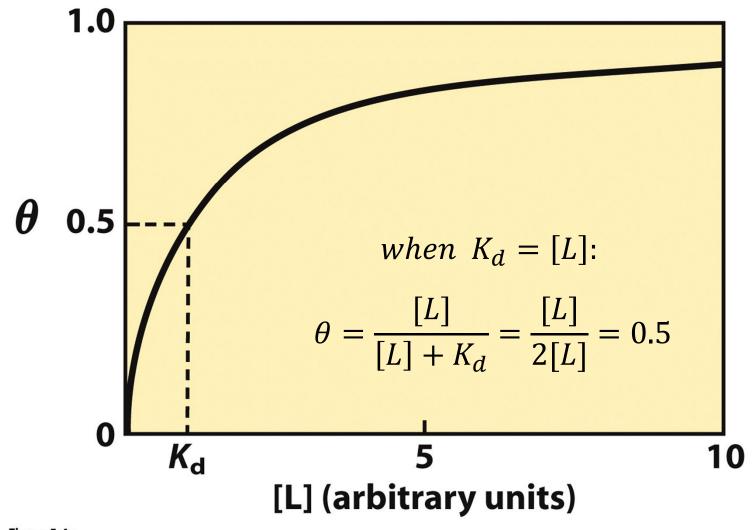
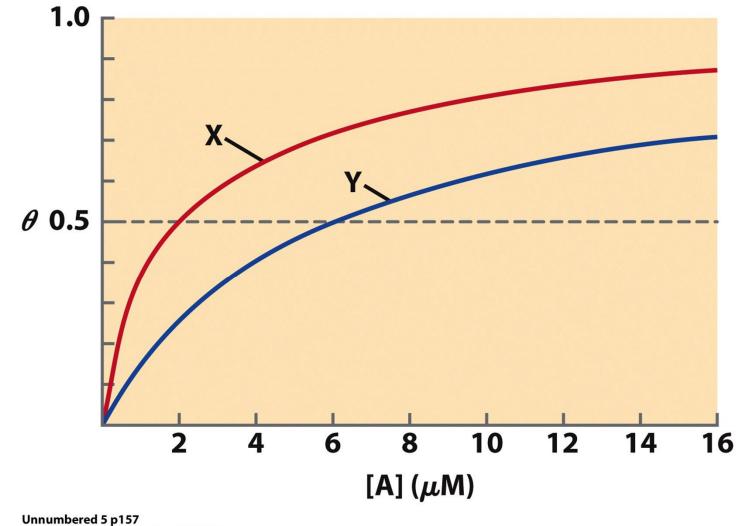


Figure 5-4a Lehninger Principles of Biochemistry, Fifth Edition © 2008 W.H. Freeman and Company

A protein with higher affinity for a ligand has a higher binding curve and lower $\rm K_{\rm d}$



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Protein-ligand dissociation constants (K_d's) vary over several orders of magnitude

TABLE 5-1	Some Protein Dissocia	tion Constants	
Protein		Ligand	<i>К_d</i> (м)*
Avidin (egg white) [†]		Biotin	1×10^{-15}
Insulin receptor (human)		Insulin	1×10^{-10}
Anti-HIV immunoglobulin (human) [‡]		gp41 (HIV-1 surface protein)	4×10^{-10}
Nickel-binding protein (E. coli)		Ni ²⁺	1 × 10 ⁻⁷
Calmodulin (rat) [§]		Ca ²⁺	$3 imes10^{-6}$
			$2 imes 10^{-5}$
	Typical	receptor-ligand interactions	
	Biotin-avidin Antibody	v-antigen Enzyme-substrate -10 10 ⁻⁸ 10 ⁻⁶ 10 ⁻⁴ 10 ⁻² <i>Iow affinity</i>	1
e 5-1		<i>K</i> _d (м)	

 Table 5-1

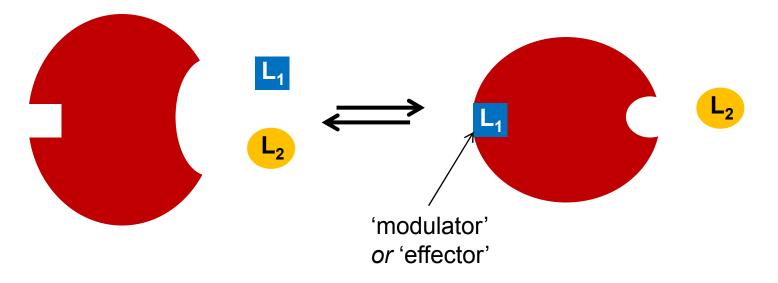
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Some proteins have varying affinity for a ligand, depending on their conformation

Allosteric protein

Binding of a ligand (L_1) to one site affects binding properties of another site (via a conformational change in the protein)

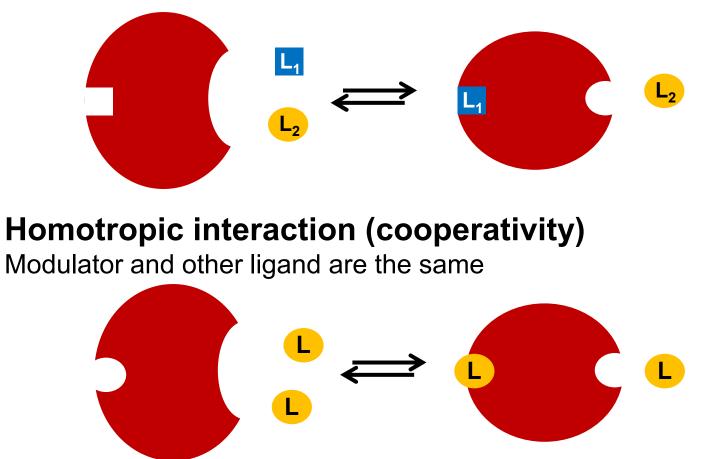


Modulator (L₁) is an 'activator' if it increases affinity at 2^{nd} site (where L₂ binds) Modulator (L₁) is an 'inhibitor' if it decreases affinity at 2^{nd} site (where L₂ binds)

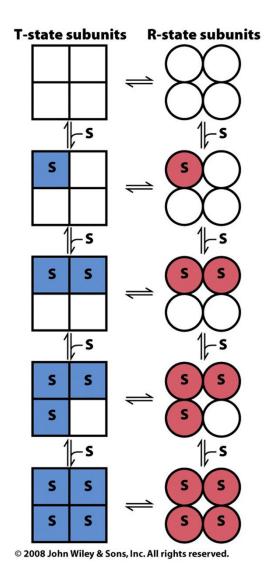
Allostery may involve different ligands, the same ligands, or both

Heterotropic interaction

Modulator and other ligand are different



The symmetry (concerted) model of cooperativity requires symmetry of the allosteric protein



- Subunits can adopt one of two possible conformations: T or R
- All subunits *must* adopt the same conformation (protein is always symmetric)
- Ligand (S) can bind to:
 - T-state with low affinity
 - R-state with high affinity
- Equilibrium between T and R states is influenced by ligand binding
- Switching between T and R is concerted; all subunits transition simultaneously

The sequential model of cooperativity allows multiple conformations for each subunit

$$= \underbrace{s}_{i} \underbrace{s}_{i}$$

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- Subunits can adopt multiple conformations
- Binding of ligand (S) induces conformational changes in the bound subunit and in neighboring subunits
- Different subunits may have different conformations, each with different ligand affinities
- Bound conformations may have higher or lower affinity for ligand than the free protein

Binding curves for allosteric proteins vary depending on the presence of modulators